Limitations of Phylogenomic Data Can Drive Inferred Speciation Rate Shifts

Jack M. Craig (b,^{1,2,3} Sudhir Kumar (b,^{*,1,2,3,4} and S. Blair Hedges (b^{*,1,2,3}

¹Center for Biodiversity, Temple University, Philadelphia, PA, USA

²Institute for Genomics and Evolutionary Medicine, Temple University, Philadelphia, PA, USA

³Department of Biology, Temple University, Philadelphia, PA, USA

⁴Center of Excellence in Genomic Medicine Research, King Abdulaziz University, Jeddah, Saudi Arabia

*Corresponding authors: E-mails: s.kumar@temple.edu; sbh@temple.edu.

Associate editor: Koichiro Tamura

Abstract

Biodiversity analyses of phylogenomic timetrees have produced many high-profile examples of shifts in the rate of speciation across the tree of life. Temporally correlated events in ecology, climate, and biogeography are frequently invoked to explain these rate shifts. In a re-examination of 15 genomic timetrees and 25 major published studies of the pattern of speciation through time, we observed an unexpected correlation between the timing of reported rate shifts and the information content of sequence alignments. Here, we show that the paucity of sequence variation and insufficient species sampling in phylogenomic data sets are the likely drivers of many inferred speciation rate shifts, rather than the proposed biological explanations. Therefore, data limitations can produce predictable but spurious signals of rate shifts even when speciation rates may be similar across taxa and time. Our results suggest that the reliable detection of speciation rate shifts requires the acquisition and assembly of long phylogenomic alignments with near-complete species sampling and accurate estimates of species richness for the clades of study.

Key words: tree of life, timetree, speciation, diversification, biodiversity.

Introduction

Speciation rates are frequently inferred in phylogenetic investigations using established computational methods to detect episodes of biodiversity evolution (Alfaro et al. 2009; Stadler 2011; Ingram and Mahler 2013; Rabosky et al. 2014; Höhna et al. 2016; Maliet et al. 2019; Paradis and Schliep 2019). Researchers have identified numerous accelerations and decelerations of speciation and proposed mechanistic drivers of biodiversity evolution across time and among clades. Consequently, the ages of inferred speciation rate shifts in many clades have been correlated with ecological interactions among species (Price et al. 2012; Claramunt and Cracraft 2015), documented changes in global or local climate (Condamine et al. 2019; Thomson et al. 2021), and earth history and biogeographical events (Quintero and Jetz 2018). Such inferences are exciting because they connect environmental and ecological factors with the rate of generation of biodiversity.

In contrast, some studies have found relatively constant rates of speciation through time, including a recent data-rich study of suboscine birds (Harvey et al. 2020), a large-scale analysis of prokaryotes (Marin et al. 2017), and a global timetree of 50,632 species (Hedges et al. 2015). Hedges et al. (2015) suggested that the major driver of speciation is the regular accumulation of genetic incompatibilities (Coyne and Orr 1989, 1998) and the stochastic nature of population isolation due to earth history events. In fact, it has been reported that rate shifts can arise spuriously even when speciation occurs at a constant rate if taxon sampling of the analyzed clade is not complete and/or the sequences analyzed are too short (Marin et al. 2017; Marin and Hedges 2018). So, although some cladespecific speciation rate shifts are expected to occur due to natural variation, it remains unclear whether many reported rate shifts are simply due to data limitations such as insufficient sequence lengths and incomplete species sampling, which are often unavoidable in biodiversity research.

In an effort to reconcile these disparate inferences of the pattern of speciation through time, which is key to understanding the drivers of speciation and biodiversity, we present results of a meta-analysis of 15 genomic timetrees and 25 published accounts of macroevolutionary rates through time at a range of phylogenetic scales.

Results

Effect of Data Richness on the Timing of Speciation Rate Shifts

To explore whether speciation rate shifts may be artifacts caused by limitations of the sequence alignments analyzed, as was found in empirical analyses (Marin et al. 2017) and simulations (Marin and Hedges 2018), we conducted an

[©] The Author(s) 2022. Published by Oxford University Press on behalf of the Society for Molecular Biology and Evolution. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https:// creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com



extensive analysis of published empirical studies. We subjected a sample of published genomic timetrees to two complementary approaches for biodiversity analysis, both of which report comparable metrics of lineage-wide macroevolutionary rates through time: TreePar (Stadler 2011) and TESS (Höhna et al. 2016). Both approaches are widely used, and their authors have tested them against comparable methods using empirical and simulated data (Stadler 2011; Höhna et al. 2016). We analyzed 15 timetrees spanning the diversity of life: plants (Pessoa-Filho et al. 2017; Ran et al. 2018; Barrera-Redondo et al. 2019), vertebrates (Kappas et al. 2016; Feng et al. 2017; Wu et al. 2017; Alfaro et al. 2018; Hughes et al. 2018; Delsuc et al. 2019; Harvey et al. 2020), and invertebrates (Blaimer et al. 2016; Fernández et al. 2018; Sann et al. 2018; Chazot et al. 2019; Kuntner et al. 2019) (table 1). They cover time depths from 45-718 My and include 11-1,939 species with alignments of 6,257-4,246,454 sites. We analyzed each timetree with TESS and TreePar and recorded the time of the inferred rate shift closest to the point at which a persistent increase in rate toward the present occurred in each analysis.

These analyses produced numerous speciation rate shifts, similar in direction and timing to those reported in rate studies of the same clades (Stadler 2011; Jetz et al. 2012; Steppan and Schenk 2017; Oliveros et al. 2019; Upham et al. 2019; Sun et al. 2020; Thomson et al. 2021). Results from TESS and TreePar analyses were highly comparable, with an average difference of $14 \pm 11\%$ in the ages of inferred rate shift and a range of 0-42% of the crown age of the clade (table 2). These patterns suggest that similar shifts in the rate of speciation are detectable by different methods both in this study and in the published literature.

We then performed linear regressions of the total number of sites against the age at which the persistent rate increase occurred in each TreePar analysis, followed by Pearson's χ^2 goodness of fit test. We observed unexpected correlations between the estimated time of speciation rate shifts and the information content of the data used in these 15 studies. The number of sites in the sequence alignment, taken as a proxy for the information content of the alignment, was inversely correlated with the age of the increase in speciation rate ($R^2 = 0.38$, P < 0.05; fig. 1*a*). That is, shorter sequence alignments produced speciation rate shifts that were more ancient, and as the alignment length increased, the inferred shift became increasingly recent. Indeed, this suggests that if we were to analyze even longer sequences, the inferred speciation rate shift would dissipate toward the present almost entirely. Thus, we can conclude that limited sequence length can bias the inferred timing of speciation rate shifts, as was shown previously (Marin et al. 2017; Marin and Hedges 2018). This correlation is not expected to result from real biological processes simply because those phenomena should have little impact on the information content of the genomic alignments we analyzed in this sample.

Interestingly, we found an even stronger correlation between overall data richness, representing the combined species count and aligned sites, and the age of speciation rate shift ($R^2 = 0.61$, $P < 10^{-3}$; fig. 1b). As with the previous correlation, the inferred speciation rate shift became more recent as the richness of the underlying data increased, reinforcing the conclusion that the shift ages are biased by the data richness used in the analysis. These two strong correlations are particularly surprising because our observed speciation rate inferences are based on data from state-of-art genomic studies in their respective clades.

Based on this result and multiple published findings from empirical and simulated work using a range of approaches (Marin et al. 2017; Marin and Hedges 2018), we then hypothesized that low information content in sequence alignments would bias the inferred pattern of speciation through time in a test case using empirical data. We tested this hypothesis by analyzing two timetrees of suboscine birds, one consisting of 881 tips assembled by a backbone-and-patch multigene approach (Jetz et al. 2012) and the other composed of 1,929 tips and inferred from a long alignment comprised over 3 million total sites (Harvey et al. 2020). The former timetree, with the addition of more species by imputation in the absence of sequence data and expanded in scope to include all birds, had previously been used to support an inferred speciation rate increase within the last 50 My (Jetz et al. 2012). By contrast, extensive analyses of the more data-rich timetree did not support major, clade-wide rate shifts, leading to the conclusion that species richness accumulates more uniformly over time (Harvey et al. 2020).

Using these two timetrees, we first examined the effect of sequence length while holding the taxonomic sampling constant by pruning each timetree to the set of 773 shared tips. Thus, both timetrees had the same set of species, but one was inferred from a much shorter alignment (site-poor) than the other (site-rich). Our analysis of the site-poor data set produced an upward shift in speciation rate at 24.6 Ma (fig. 1c), whereas that of the site-rich data set produced a 53% younger rate shift at 11.7 Ma (fig. 1d). This pattern is concordant with that shown in figure 1a and b, allowing us to predict that the inclusion of all 1,929 species in the large site-rich data set would further reduce the bias in the inferred speciation rate shift. Indeed, a much younger rate shift was observed at 4.5 Ma in this analysis, an 81% decrease from 24.6 Ma (fig. 1e). Therefore, the bias in the inferred speciation rate shifts is predictable based on the known effects of site- and species-deficits (Marin et al. 2017; Marin and Hedges 2018).

This result suggests that the rate shift observed in the sitepoor data sets may be explained by a statistical bias stemming from low information content of the underlying sequence alignment. This explanation would obviate the need to invoke myriad biological drivers to explain the observed shifts in speciation rates.

The Effect of Taxonomic Coverage on the Pattern of Speciation through Time

To assess the effect of incomplete taxonomic sampling, we tested two scenarios. In one, taxonomic knowledge is highly complete, meaning that extant biodiversity is well characterized and estimates of taxonomic sampling are highly accurate. In the other, taxonomic knowledge is poor, meaning that there remain many undescribed species which precludes accurate estimation of taxonomic sampling. For both cases, we

Reference	Journal	Root Age	Species Present	Number of Sites	
Alfaro et al. (2018)	Nature Ecology and Evolution	139.57	118	302,488	
Barrera-Redondo et al. (2019)	Molecular Plant	103.11	11	228,983	
Blaimer et al. (2016)	Molecular Phylogenetics and Evolution	46.52	25	733,400	
Chazot et al. (2019)	Systematic Biology	108.74	994	6,257	
Delsuc et al. (2019)	Current Biology	64.85	40	15,157	
Feng et al. (2017)	Proceedings of the National Academy of Sciences	427.00	164	88,302	
Fernández et al. (2018)	Current Biology	470.16	168	1,871,676	
Harvey et al. (2020)	Science	44.73	1,939	3,708,449	
Hughes et al. (2018)	Proceedings of the National Academy of Sciences	441.00	305	555,288	
Kappas et al. (2016)	PLoS One	141.30	66	15,557	
Kuntner et al. (2019)	Systematic Biology	181.68	34	89,212	
Pessoa-Filho et al. (2017)	BMC Genomics	54.10	30	138,976	
Ran et al. (2018)	Molecular Phylogenetics and Evolution	718.91	16	4,246,454	
Sann et al. (2018)	BMC Evolutionary Biology	242.06	185	283,008	
Wu et al. (2017)	Current Biology	262.80	45	872,511	
	Minimum	44.73	11.00	6,257.00	
	Maximum	718.91	1,939.00	4,246,454.00	

Table 1. The Fifteen Genomic Timetrees Used	in Our Regressions	5.
---	--------------------	----

 Table 2. Age of Speciation Rate Shifts Predicted from 15 Phylogenomic Timetrees.

Group	Species Count	Root Age (Ma)	TreePar Shift (Ma)	TESS Shift (Ma)
Suboscine birds	1,939	44.7	4.47	7.77
Butterflies	994	108.7	36.32	53.73
Fishes	305	441.0	99.67	99.47
Bees	185	242.1	80.85	134.57
Spiders	168	470.1	131.65	263.11
Frogs	164	427.0	165.68	145.80
Fishes	118	139.6	61.69	79.39
Catfishes	66	141.3	98.06	101.32
Placentals	45	262.8	97.23	133.45
Sloths	40	64.9	27.50	37.90
Spiders	34	181.7	44.33	32.01
Grasses	30	54.1	24.89	29.42
Ants	25	46.5	13.86	33.46
Pines	16	718.9	188.36	335.76
Cucumbers	11	103.1	34.44	31.66

Note.—Age of shift times are from the maximum likelihood analysis in TreePar (Stadler 2011) and Bayesian analysis in TESS (Höhna et al. 2016). The ages represent discrete times at which a persistent upward rate shift toward the present was observed. TESS rate shift is approximate because results are reported as a distribution of shift times. Ma, million years ago.

performed a series of TESS analyses on 75%, 50%, 25%, and 5% of the tips randomly selected from empirical timetrees and noted the point at which the inferred speciation pattern changed. Importantly, in the case of excellent taxonomic knowledge, we were able to supply TESS with an accurate estimate of sampling fraction (the percentage of known species richness represented, ρ), but not in the case of poor taxonomic knowledge.

For the scenario in which taxonomic knowledge is high, we used the data-rich phylogeny of suboscine birds (Harvey et al. 2020). This is an uncommonly well-studied group, as reflected in the 670 research items on Google Scholar including the term "Tyranni" in the year 2021 alone. This allowed us to generate accurate values of ρ for TESS that can better

compensate for incomplete sampling (Höhna et al. 2011). We observed a nearly constant rate of speciation through time at all levels of taxonomic coverage (fig. 2*a*). This suggests that when taxonomic knowledge is high and ρ can be parameterized accurately, the overall pattern of speciation, including shifts in rate, is robust to changes in ρ .

Taxonomic knowledge is rarely complete, however, especially in large clades where tens or hundreds of species are yet to be described. For the scenario in which the real species richness is poorly understood or poorly represented in the timetree (or both), we used the timetrees of amphibians (4,179 tips) and squamate reptiles (6,291 tips) derived from the March 2021 alpha release of the *TimeTree* database (Kumar et al. 2017), which differs minimally from the fifth



Fig. 1. An unexpected relationship between the age of speciation rate shift and the number of aligned sites and overall data richness. (*a*) Regression of the number of aligned sites (log-scale) against the time of the rate shift reported from TreePar analyses of 15 site-rich timetrees. The gray line shows the log-linear fit (slope = -0.14, $R^2 = 0.38$; P < 0.02). (*b*) Regression of overall data richness (log(sites × species)) against the time of the rate shift using TreePar analyses of 15 site-rich timetree. The gray line shows the log-linear fit (slope = -0.14; $R^2 = 0.61$, P < 0.001). The pattern of speciation inferred by TESS for the (*c*) site-poor and (*d*) site-rich timetree of suboscine birds pruned to include only shared tips between two studies (773 species). (*e*) The pattern of speciation inferred by TESS for the full site-rich timetree. In panels (*c*-*e*) blue lines mark the time at which a TreePar analysis of the same timetree reported a persistent upward rate shift toward the present. We used a TreePar model with seven rate shifts because it fit the data best and gave us the most resolution. These shift times (blue) are largely concordant with those recovered from TESS (grey).



Fig. 2. Tests reveal very little difference in the pattern of speciation in a scenario where taxonomic knowledge is sufficient to make confident estimates of coverage. (*a*) Speciation rates inferred from the TESS analyses of the data-rich timetree (Harvey et al. 2020) at five different levels of sampling. Darker shading is proportional to greater ρ at the following levels: 100% (darkest blue), 75%, 50%, 25%, and 5% of the total timetree. Note that the overall pattern of speciation is visually similar across all levels. (*b*) Regression of ρ against the speciation rate hyperprior inferred from TESS analysis.

edition. Given that these data represent subsets of an already taxonomically incomplete data set, and that the large clades we studied likely contain hundreds of undescribed species, further complicating estimates of coverage, we did not provide a ρ value in these analyses. Instead, we used these

timetrees as representatives of large-scale but poorly sampled timetrees for which ρ is difficult to estimate reliably, and unavoidable result of ongoing taxonomic and systematic work. For both clades, the patterns of speciation we found in our analyses of the 75%-sampled timetrees differed



Fig. 3. Tests reveal differences in the pattern of speciation and a correlation between species-rich data and age of speciation rate shifts in a scenario where limited taxonomic knowledge precludes accurate estimates of coverage. (*a*, *b*) Speciation rates inferred from TESS analyses of the (*a*) amphibian and (*b*) squamate reptile timetrees at five different levels of sampling. Darker shading is proportional to greater ρ at the following levels: 100% (darkest blue), 75%, 50%, 25%, and 5% of the total timetree. Note that the speciation rate plots for the 75% and 100%-sampled trees are visually similar, but the pattern changes below 50%. (*c*, *d*) Regressions of ρ against the speciation rate hyperprior inferred from TESS analyses for the (*c*) amphibian and (*d*) squamate reptile timetrees.

minimally from those of the 100%-sampled timetrees, whereas those of the 50% timetrees and below were clearly different (fig. 3*a* and *b*). This suggests that $\rho = 75\%$ is a conservative minimum threshold when the real species richness is poorly characterized or poorly represented in the timetree.

In these analyses, we also observed taxonomic coverage to impact the inferred hyperprior of speciation rate. For the case of excellent taxonomic knowledge (suboscine birds), we found ρ to be positively correlated with the inferred hyperprior rate of speciation (polynomial regression $R^2 = 0.98$, P < 0.05). Minimum and maximum inferred rates differed by 0.16, 39% of the maximum inferred rate (fig. 2b). For the case of poor taxonomic knowledge (amphibians and squamate reptiles), we found ρ to be inversely correlated with the inferred hyperprior rate of speciation ($R^2 = 1.00$, $P < 10^{-5}$ for amphibians, $R^2 = 1.00$, $P < 10^{-5}$ for squamate reptiles). Minimum and maximum inferred rates differed by 0.032 for amphibians, 61% of their maximum rate, and 0.028 for squamate reptiles, 55% of their maximum rate (fig. 3c and d). Interestingly, the direction of these correlations is reversed when compared with the data-rich scenario and the difference between minimum and maximum rates increased, despite the same percentages of each timetree being sampled in each scenario. This phenomenon provides further evidence that poor taxonomic knowledge strongly influences speciation rate analyses.

Taken together, these results suggest that inferences of the instantaneous or prior rate of speciation for any group are

likely biased by the number of tips included in the analysis, making it impossible to compare speciation rate directly between clades of different scales even given high taxonomic knowledge. However, the pattern of speciation through time, especially when taxonomic knowledge is excellent or at least most known species are included, is comparable.

The Effect of Scale on the Pattern of Speciation through Time

Although our results suggest that the number of aligned sites and taxonomic sampling of a given timetree may influence its inferred pattern of speciation, it is possible that these patterns may instead be attributed to the effect of taxonomic scale. So, to determine the influence of clade size, we performed the TESS analysis on timetrees of the seven largest suboscine bird families extracted from the data-rich timetree (Harvey et al. 2020). They varied in taxonomic scale from 21 to 542 tips (compared with 1,939 for the complete timetree) but were otherwise identical in all aspects of tree building (fig. 4). We found that several families were characterized by roughly constant rates of speciation through time, like the complete timetree. This included both the largest and smallest families, the Tyrannidae (542 tips; fig. 4j) and the Formicariidae (21 tips; fig. 4c), suggesting that the pattern of rate constancy was not strongly influenced by taxonomic scale. By contrast, some families appeared to have undergone shifts in rate, such as the Cotingidae (79 tips; fig. 4b) and Rhinocryptidae (80 tips;



FIG. 4. Inferred patterns of speciation rate through time among complete timetrees of the seven largest families of suboscine birds. All timetrees are derived from Harvey et al. (2020), and therefore share an underlying sampling fraction and genomic alignment. (*a*) Full timetree, (*b*) Cotingidae (79 tips), (*c*) Formicariidae (21 tips), (*d*) Furnariidae (381 tips), (*e*) Grallariidae (66 tips), (*f*) Pipridae (61 tips), (*g*) Rhinocryptidae (80 tips), (*h*) Thamnophilidae (356 tips), (*i*) Tityridae (37 tips), (*j*) Tyrannidae (542 tips).

fig. 4g). This may be due to unevenness in taxonomic knowledge (more undescribed species in some families than others) or differences in sequence information content in some taxa or regions. More compellingly, it may reflect real biological processes that are not detectable at higher taxonomic scales when clade-specific patterns are combined. For example, some lineages likely speciate more or less rapidly than others due to environmental pressures, but when assessed as part of a larger-scale analysis, this variation is obscured, resulting in an overall constant rate. This is an important limitation of analyses of speciation rate through time that can only be addressed by more data-rich timetrees and finer-scale analyses.

To further assess the effect of scale on inferences of speciation rate, this time controlling for the clade of study, we compared the above timetrees of Tyrannidae (542 tips) and Tityridae (37 tips), both of which are characterized by comparable patterns of mostly constant speciation through time. We then randomly pruned the Tyrannidae timetree to 37 tips and performed the same TESS analysis, including an accurate estimate of sampling fraction. We still found a constant rate of speciation in this case as in the complete Tyrannidae and Tityridae timetrees (fig. 5).

All these results suggest that TESS is capable of accounting for taxonomic scale even when the timetrees analyzed are not taxonomically complete. Thus the pattern of rate constancy we observe cannot exclusively be explained by taxonomic scale, as we found repeatable results in both large and small clades as well as small subsets of larger clades. The terminal drop in the speciation rate in several clades (e.g., fig. 4*a*) is a common taxonomic artifact and ignored here when comparing the three analyses.

Data Richness Thresholds for Speciation Rate Analysis Based on the empirical results presented here and simulations published elsewhere (Marin et al. 2017; Marin and Hedges 2018), we created a schematic to capture relevant site- and species-richness thresholds needed to reliably detect a uniform rate of speciation without false-positive inferences of rate shifts (fig. 6). The vertical axis represents the taxonomic coverage of the timetree under study. We assume an imperfect knowledge of the real species richness, which precludes reliable estimates of sampling fraction, as is unavoidably the case in many empirical data sets, especially for very large groups. If sampling fraction can be estimated reliably, taxonomic coverage less than 75% can still yield reliable results, but we caution that this may be difficult to achieve. These threshold values were derived from our tests of sampling fraction (figs. 2 and 3) and published discussions of the "taxonomic artifact" (Hedges et al. 2015, Marin and Hedges 2018). The horizontal axis represents the number of variable sites present in the alignment used in tree building divided by the total known species richness of the clade. Based on previous work (Marin and Hedges 2018), more sites are needed when the extinction rate (μ) is high, or when taking total aligned sites as a proxy for variable sites, which is a useful convention in comparative projects. This threshold was derived from the simulation work of Marin and Hedges (2018), who characterized the lack of adequate variable sites as "the small sample artifact."

Thus, for a timetree to yield reliable inferences of the pattern of speciation through time, accounting for gaps in taxonomic knowledge and for the statistical biases we report here, we propose a conservative minimum of 75 aligned sites



FIG. 5. Tests reveal a constant rate of speciation inferred across multiple taxonomic scales, even in taxonomically incomplete timetrees. A roughly constant pattern of speciation is seen in: (*a*) the full timetree of Tyrannidae (542 tips), (*b*) the full timetree of Tityridae (37 tips), and (*c*) the timetree of Tyrannidae pruned to the same taxonomic scale as that of Tityridae (37 tips).

for every described species in the clade of study, and 75% taxonomic sampling of all known species. Improvements in taxonomic knowledge may allow for precise estimates of sampling fraction and less complete sampling, but it is prudent to not make too many assumptions about known species richness.

Note that, although it is impossible to sample extinct lineages using timetrees of extant species, meaning that no analysis could ever include all species ever to have existed, the extinction of these lineages is part of the birth-death process that produced the timetree. Thus, a complete timetree of extant species represents a snapshot in an ongoing process of diversification. The promise of modeling approaches like TESS and TreePar is to use the timetree to infer rates of speciation and extinction through time by modeling this birth-death process, and therefore missing extinct species are appropriately modeled.



FIG. 6. Thresholds of data richness needed to infer speciation rates reliably. The horizontal axis represents genomic variability (ratio of variable sites/number of species). The vertical axis represents taxonomic completeness (species sampling fraction of the timetree). Numbered dots represent the studies examined for which the data were available and compatible with this schematic, with numbers following table 3 for all studies which could be plotted.

Discussion

The results presented here demonstrate that insufficient data richness, either in the alignment used in tree building or in the number of species included, can bias inferences of the pattern of speciation through time in predictable ways. Based on these findings, we next aimed to determine the impact these artifacts have on the field. We conducted a literature review to identify published macroevolutionary rate analyses and assessed whether they met the thresholds we suggest to avoid inferring spurious rate shifts (fig. 6). Of 25 recent, high-profile studies reporting major biological drivers of rate shifts, median sampling fraction (the percentage of known species richness represented, ρ), was 28%, below the conservative threshold of \sim 75% we determined empirically for understudied groups. More problematically, the median of 7.70 total aligned sites per known species is also well below our conservative threshold of 75 where the number of sites is taken as a proxy for the information content of the alignment.

Unfortunately, 24 of the 25 studies analyzed did not meet one or both of the criteria for reliable rate estimates. Only one study (Harvey et al. 2020), which we analyzed earlier in this article, exceeds the required thresholds. Its analysis does not suggest any ancient rate shifts, consistent with a relatively constant global rate hypothesis proposed in previous timetree analyses (Hedges et al. 2015; Marin et al. 2017). These patterns reveal that many published analyses of speciation rate risk finding false-positive signals of rate shifts simply due to unavoidable limitations in their data collection, such as specimen availability and lab resources.

However, we consider two alternative explanations for the patterns we document here. First, the studies we investigated used a variety of tree building and timing approaches which may affect their inferences. For example, the Yule branching process prior implemented in BEAST (Bouckaert et al. 2014) has been shown to yield timetrees with significantly older dates for shallow nodes than timetrees built using a birthdeath prior, even given the same fossil calibrations (Condamine et al. 2015). This, in turn, has an impact on subsequent analyses of speciation rate. Further, shorter alignments may not be equally informative at different depths in the timetree based on the choice of genes included. If deeper nodes are characterized by less phylogenetically informative genetic variation, for example due to saturation, they will be more strongly influenced by a problematic tree prior. However, the false-positive rate shifts we document here cannot be simply ascribed to poorly fitting tree priors in Bayesian dating analyses because similar trends have been observed

Authors	Year	PMID	ID in <mark>fig. 6</mark>	Species Present	Total Species Richness	Sampling Fraction	Total Sites	Sites/Species
Thomson et al.	2021	33558231	1	279	348	0.80	13,659	39.25
McCord et al.	2021	34705840	2	330	422	0.78	8,238	19.52
Sun et al.	2020	32620894	3	19,740	70,000	0.28	7,900	0.11
Han et al.	2020	31394010	4	NA	NA	NA	NA	NA
Harvey et al.	2020	33303617	5	1,939	1,939	1.00	3,708,449	1,912.56
De-Silva et al.	2019	27546953	6	54	193	0.28	NA	NA
Folk et al.	2019	31085636	7	627	4,823	0.13	324,662	67.32
Oliveros et al.	2019	30936315	8	137	10,698	0.01	2,464,926	230.41
Condamine et al.	2019	31486279	9	NA	NA	NA	NA	NA
Upham et al.	2019	NA	10	5,911	5,911	1.00	39,099	6.61
Sayol et al.	2019	31518002	11	9,993	17,504	0.57	NA	NA
Xue et al.	2019	31639525	12	835	2,386	0.35	11,211	4.70
Liu et al.	2018	29550535	13	89	345	0.26	7,175	20.80
Castro-Insua et al.	2018	29884843	14	165	12,661	0.01	35,603	2.81
Shao and Li	2018	29803949	15	91	20,574	0.00	1,500	0.07
Wang et al.	2018	30381380	16	285	491	0.58	7,080	14.42
Murray et al.	2018	30429246	17	70	10,000	0.01	284,607	28.46
Seeholzer et al.	2017	28071791	18	284	301	0.94	NA	NA
Steppan and Schenk	2017	28813483	19	900	1,620	0.56	3,070	1.90
Roalson and Roberts	2016	26880147	20	786	3,300	0.24	29,000	8.79
Claramunt and Cracraft	2015	26824065	21	230	17,504	0.01	NA	NA
Baker and Couvreur	2013	NA	22	125	2,105	0.06	NA	NA
Hou et al.	2011	21844362	23	114	1,012	0.11	5,088	5.03
Roelants et al.	2007	17213318	24	171	11,712	0.01	3,750	0.32
Hughes and Eastwood	2006	16801546	25	98	256	0.38	1,000	3.91
Median				279	2,386	0.28	9,725	7.70

Table 3. The 25 Macroevolutionary Rate Analyses Used in Our Literature Review.

Note.—NAs indicate studies which either did not provide these data or summary statistics in their publication, supplementary material, Supplementary Material online, or in a readily available online database, or used a timetree reconstruction method incompatible with our analyses.

(Marin et al. 2017) in timetrees inferred using the RelTime approach (Tamura et al. 2012) which does not use a speciation tree prior. We recommend researchers consider such an approach to corroborate their results in future work.

Second, we report a pronounced terminal (<5 My) decline in speciation in many of our analyses even when taxonomic coverage is high (e.g., fig. 1e), which is also evident in many published analyses. This is not exclusively due to the biases we discuss here, but is likely a result of reticulate evolution among incompletely diverged populations below the species level, which violates the assumption of branching evolution on which many methods rely (Hedges et al. 2015). For example, TESS models diversification as a birth–death process, but among subspecific populations "birth" has not taken place which may result in an inferred decline in speciation (Hedges et al. 2015). Therefore, an abrupt terminal decline in rate may be inferred even from very data-rich timetrees, though the pattern is likely to be more pronounced when taxonomic sampling is less complete or difficult to estimate accurately. In conclusion, researchers studying the pattern of diversification through time need to carefully consider the potential limitations of their data. We propose that inferences of major speciation rate shifts still await large-scale, site-rich data sets with taxonomic coverage as close to the known species richness of the clade of study as possible. Fortunately, we see promise for such data sets arising from growing coalitions to assemble comprehensive species collections, increasing affordability and accuracy of genome sequencing, and advances in the availability of taxonomic, systematic, and bioinformatic data sharing (Kumar et al. 2017; Schoch et al. 2020).

Materials and Methods

Timetrees Used in Analyses

We used a variety of published timetrees in our analyses. For the regressions of data richness against rate shift time, we used a sample of 15 published genomic timetrees, all of which made their full alignments available (table 2). To compare site-poor and site-rich timetrees of suboscine birds, we used the timetree of birds freely distributed at https://birdtree.org/ as our site-poor timetree. We used their timetree built from only species for which genetic data were available, with no tips added by imputation, following the Ericson backbone taxonomy (Ericson et al. 2006). Our site-rich timetree was the nearly completely sampled timetree of suboscine birds available at www.mgharvey.org. We used the complete, strictly filtered (HGAPF) tree based on the Clements taxonomy (Clements 2008) and treated sampling fraction (ρ) as 1.00. For our literature review, we assembled a collection of 25 published analyses of macroevolutionary rates (table 3). These were not selected based on their underlying data richness or on whether they found rate shifts, only on whether they investigated macroevolutionary rates through time and made their tree files readily available. For the tests of ρ in amphibians and squamate reptiles, we used timetrees derived from the March 2021 alpha release of the fifth edition of TimeTree (Kumar et al. 2017).

Bayesian Analyses in TESS

TESS (Höhna et al. 2016) is software package which implements a Bayesian approach in R based on the reconstructed evolutionary process of Nee et al. (1994), modified by Höhna (2015) to incorporate rate shifts through time. It uses an adaptive MCMC process (Haario et al. 1999) to estimate the number of rate shifts, their ages, and their magnitudes across a timetree. For all tests, we ran the CoMET analysis in TESS for 20,000 iterations, the first 1,000 of which were discarded as burn-in, leaving other parameters at default values. We also used TESS to generate empirical hyperpriors based on the input timetree. TESS accounts for incompletely sampled timetrees by incorporating a user-provided value of taxonomic coverage (the probability of a known lineages appearing in the timetree, or sampling fraction, ρ), allowing the overall patterns of inferred rate to remain comparable across a range of sampling regimes (Höhna et al. 2011, 2016). We implemented this parameter differently in each test, as discussed above.

Maximum Likelihood Analyses in TreePar

TreePar (Stadler 2011) is a is software package which implements a maximum likelihood approach based on a variation of the simple birth-death model incorporating a given number of rate shifts, then tests the improvement in model fit contributed by successive additional rate shifts. We used TreePar to estimate speciation rates through time and the most likely number of rate shifts in each timetree. Following Hedges et al. (2015), we tested eight models, from zero rate shifts (constant rate) to seven. TreePar allows the user to provide a value of ρ at each time-slice but given that accurate estimates of sampling fraction are difficult even in the present, and likely impossible when accounting for extinct lineages at ancient time points, we declined to provide this parameter in our analyses for consistency between analyses and to minimize error. For all tests, we estimated rates and allowed shifts at 50 intervals divided equally over the age of each timetree, ignoring the first one percent of the total age, where the low number of extant lineages present makes inferences of rates

unreliable (Hedges et al. 2015). We also discarded the youngest ten percent of the age of each timetree, where subspecific evolution is predominantly reticulate, not branching, which precludes analysis by most models (Hedges et al. 2015). Given that TreePar only allows a set number of rate shifts, we aimed to avoid the risk of finding a false-positive rate shift in these problematic regions. We then compared these eight successive rate shift models by LRT to select the best model at the 95% confidence level.

Supplementary Material

Supplementary data are available at *Molecular Biology and Evolution* online.

Acknowledgments

The authors gratefully acknowledge Jose Barba-Montoya, Michael G. Harvey, Qiqing Tao, Sudip Sharma, Michael Suleski, and Tanja Stadler for their assistance. This work was supported by grants from the U.S. National Science Foundation to S.B.H. and S.K. (DBI 1932765), National Institutes of Health to S.K. (GM0126567-02), and Temple University.

References

- Alfaro ME, Faircloth BC, Harrington RC, Sorenson L, Friedman M, Thacker CE, Oliveros CH, Černý D, Near TJ. 2018. Explosive diversification of marine fishes at the Cretaceous-Palaeogene boundary. *Nat Ecol Evol.* 2(4):688–696.
- Alfaro ME, Santini F, Brock C, Alamillo H, Dornburg A, Rabosky DL, Carnevale G, Harmon LJ. 2009. Nine exceptional radiations plus high turnover explain species diversity in jawed vertebrates. *Proc Natl Acad Sci U S A*. 106(32):13410–13414.
- Barrera-Redondo J, Ibarra-Laclette E, Vázquez-Lobo A, Gutiérrez-Guerrero YT, Sánchez de la Vega G, Piñero D, Montes-Hernández S, Lira-Saade R, Eguiarte LE. 2019. The genome of *Cucurbita argyrosperma* (silver-seed gourd) reveals faster rates of protein-coding gene and long noncoding RNA turnover and neofunctionalization within *Cucurbita. Mol Plant.* 12(4):506–520.
- Blaimer BB, LaPolla JS, Branstetter MG, Lloyd MW, Brady SG. 2016. Phylogenomics, biogeography and diversification of obligate mealybug-tending ants in the genus Acropyga. *Mol Phylogenet Evol.* 102:20–29.
- Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu CH, Xie D, Suchard MA, Rambaut A, Drummond AJ. 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. PLoS Comput Biol. 10:1–6.
- Chazot N, Wahlberg N, Freitas AVL, Mitter C, Labandeira C, Sohn JC, Sahoo RK, Seraphim N, De Jong R, Heikkilä M. 2019. Priors and posteriors in Bayesian timing of divergence analyses: the age of butterflies revisited. Syst Biol. 68(5):797–813.
- Claramunt S, Cracraft J. 2015. A new time tree reveals Earth history's imprint on the evolution of modern birds. Sci Adv. 1(11):1–13.
- Clements J. 2008. The Clements checklist of birds of the world. 6th ed. Ithaca (NY): Comstock Publishing Associates.
- Condamine FL, Nagalingum NS, Marshall CR, Morlon H. 2015. Origin and diversification of living cycads: a cautionary tale on the impact of the branching process prior in Bayesian molecular dating. BMC Evol Biol. 15:1–18.
- Condamine FL, Rolland J, Morlon H. 2019. Assessing the causes of diversification slowdowns: temperature-dependent and diversitydependent models receive equivalent support. *Ecol Lett.* 22(11):1900–1912.
- Coyne JA, Orr HA. 1989. Patterns of speciation in Drosophila. *Evolution* 43(2):362-381.

- Delsuc F, Kuch M, Gibb GC, Karpinski E, Hackenberger D, Szpak P, Martínez JG, Mead JI, McDonald HG, MacPhee RDE, et al. 2019. Ancient mitogenomes reveal the evolutionary history and biogeography of sloths. *Curr Biol.* 29(12):2031–2042.e6.
- Ericson PGP, Anderson CL, Britton T, Elzanowski A, Johansson US, Källersjö M, Ohlson JI, Parsons TJ, Zuccon D, Mayr G. 2006. Diversification of Neoaves: integration of molecular sequence data and fossils. *Biol Lett.* 2(4):543–547.
- Feng YJ, Blackburn DC, Liang D, Hillis DM, Wake DB, Cannatella DC, Zhang P. 2017. Phylogenomics reveals rapid, simultaneous diversification of three major clades of Gondwanan frogs at the Cretaceous– Paleogene boundary. Proc Natl Acad Sci U S A. 114(29):E5864–E5870.
- Fernández R, Kallal RJ, Dimitrov D, Ballesteros JA, Arnedo MA, Giribet G, Hormiga G. 2018. Phylogenomics, diversification dynamics, and comparative transcriptomics across the spider tree of life. *Curr Biol.* 28(9):1489–1497.e5.
- Haario H, Saksman E, Tamminen J. 1999. Adaptive proposal distribution for random walk Metropolis algorithm. *Comput Stat.* 14(3):375–395.
- Harvey MG, Bravo GA, Claramunt S, Cuervo AM, Derryberry GE, Battilana J, Seeholzer GF, McKay JS, O'Meara BC, Faircloth BC, et al. 2020. The evolution of a tropical biodiversity hotspot. *Science* 370(6522):1343–1348.
- Hedges SB, Marin J, Suleski M, Paymer M, Kumar S. 2015. Tree of life reveals clock-like speciation and diversification. *Mol Biol Evol.* 32(4):835–845.
- Höhna S. 2015. The time-dependent reconstructed evolutionary process with a key-role for mass-extinction events. J Theor Biol. 380:321-331.
- Höhna S, May MR, Moore BR. 2016. TESS: an R package for efficiently simulating phylogenetic trees and performing Bayesian inference of lineage diversification rates. *Bioinformatics* 32(5):789–791.
- Höhna S, Stadler T, Ronquist F, Britton T. 2011. Inferring speciation and extinction rates under different sampling schemes. *Mol Biol Evol.* 28(9):2577–2589.
- Hughes LC, Ortí G, Huang Y, Sun Y, Baldwin CC, Thompson AW, Arcila D, Betancur R, Li C, Becker L, et al. 2018. Comprehensive phylogeny of ray-finned fishes (Actinopterygii) based on transcriptomic and genomic data. *Proc Natl Acad Sci U S A*. 115(24):6249–6254.
- Ingram T, Mahler DL 2013. SURFACE: detecting convergent evolution from comparative data by fitting Ornstein-Uhlenbeck models with stepwise Akaike Information Criterion. *Methods Ecol Evol*. 4(5):416–425.
- Jetz W, Thomas GH, Joy JB, Hartmann K, Mooers AO. 2012. The global diversity of birds in space and time. *Nature* 491(7424):444–448.
- Kappas I, Vittas S, Pantzartzi CN, Drosopoulou E, Scouras ZG. 2016. A time-calibrated mitogenome phylogeny of catfish (Teleostei: Siluriformes). PLoS One 11(12):e0166988.
- Kumar S, Stecher G, Suleski M, Hedges SB. 2017. TimeTree: a resource for timelines, timetrees, and divergence times. *Mol Biol Evol.* 34(7):1812–1819.
- Kuntner M, Hamilton CA, Cheng RC, Gregorič M, Lupše N, Lokovšek T, Lemmon EM, Lemmon AR, Agnarsson I, Coddington JA, et al. 2019. Golden orbweavers ignore biological rules: phylogenomic and comparative analyses unravel a complex evolution of sexual size dimorphism. Syst Biol. 68(4):555–572.
- Maliet O, Hartig F, Morlon H. 2019. A model with many small shifts for estimating species-specific diversification rates. *Nat Ecol Evol.* 3(7):1086–1092.

- Marin J, Battistuzzi FU, Brown AC, Hedges SB. 2017. The timetree of prokaryotes: new insights into their evolution and speciation. *Mol Biol Evol.* 34(2):437–446.
- Marin J, Hedges SB. 2018. Undersampling genomes has biased time and rate estimates throughout the tree of life. *Mol Biol Evol.* 35(8):2077–2084.
- Nee S, May RM, Harvey PH. 1994. The reconstructed evolutionary process. Philos Trans R Soc Lond B Biol Sci. 344(1309):305-311.
- Oliveros CH, Field DJ, Ksepka DT, Keith Barker F, Aleixo A, Andersen MJ, Alström P, Benz BW, Braun EL, Braun MJ, et al. 2019. Earth history and the passerine superradiation. *Proc Natl Acad Sci U S A*. 116(16):7916–7925.
- Paradis E, Schliep K. 2019. Ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* 35(3):526–528.
- Pessoa-Filho M, Martins AM, Ferreira ME. 2017. Molecular dating of phylogenetic divergence between Urochloa species based on complete chloroplast genomes. *BMC Genomics* 18(1):14.
- Price SA, Hopkins SSB, Smith KK, Roth VL. 2012. Tempo of trophic evolution and its impact on mammalian diversification. *Proc Natl Acad Sci U S A*. 109(18):7008–7012.
- Quintero I, Jetz W. 2018. Global elevational diversity and diversification of birds. *Nature* 555(7695):246–250.
- Rabosky DL, Grundler M, Anderson C, Title P, Shi JJ, Brown JW, Huang H, Larson JG. 2014. BAMMtools: an R package for the analysis of evolutionary dynamics on phylogenetic trees. *Methods Ecol Evol.* 5(7):701–707.
- Ran JH, Shen TT, Wu H, Gong X, Wang XQ. 2018. Phylogeny and evolutionary history of Pinaceae updated by transcriptomic analysis. *Mol Phylogenet Evol.* 129:106–116.
- Sann M, Niehuis O, Peters RS, Mayer C, Kozlov A, Podsiadlowski L, Bank S, Meusemann K, Misof B, Bleidorn C, et al. 2018. Phylogenomic analysis of Apoidea sheds new light on the sister group of bees. BMC Evol Biol. 18:1–15.
- Schoch CL, Ciufo S, Domrachev M, Hotton CL, Kannan S, Khovanskaya R, Leipe D, McVeigh R, O'Neill K, Robbertse B, et al. 2020. NCBI Taxonomy: a comprehensive update on curation, resources and tools. *Database* 2020:1–21.
- Stadler T. 2011. Mammalian phylogeny reveals recent diversification rate shifts. *Proc Natl Acad Sci U S A*. 108(15):6187–6192.
- Steppan SJ, Schenk JJ. 2017. Muroid rodent phylogenetics: 900-Species tree reveals increasing diversification rates. *PLoS One* 12(8):e0183070.
- Sun M, Folk RA, Gitzendanner MA, Soltis PS, Chen Z, Soltis DE, Guralnick RP. 2020. Recent accelerated diversification in rosids occurred outside the tropics. *Nat Commun.* 11:1–12.
- Tamura K, Battistuzzi FU, Billing-Ross P, Murillo O, Filipski A, Kumar S. 2012. Estimating divergence times in large molecular phylogenies. *Proc Natl Acad Sci U S A*. 109(47):19333–19338.
- Thomson RC, Spinks PQ, Bradley Shaffer H. 2021. A global phylogeny of turtles reveals a burst of climate-associated diversification on continental margins. *Proc Natl Acad Sci U S A*. 118(7):e2012215118.
- Upham NS, Esselstyn JA, Jetz W. 2019. Inferring the mammal tree: species-level sets of phylogenies for questions in ecology, evolution, and conservation. *PLoS Biol.* 17(12):e3000494.
- Wu J, Yonezawa T, Kishino H. 2017. Rates of molecular evolution suggest natural history of life history traits and a post-K-Pg nocturnal bottleneck of placentals. *Curr Biol.* 27(19):3025–3033.e5.